

TransIT-X2[®] Dynamic Delivery System for CRISPR/Cas9 Ribonucleoprotein (RNP) + DNA Oligo (ssODN) Delivery

Instructions for use with MIR 6000, 6003, 6004, 6005, 6006, 6010



SPECIFICATIONS

Storage	Store <i>TransIT-X2[®] Dynamic Delivery System</i> tightly capped at -20°C . Before each use , warm to room temperature and vortex gently.
Product Guarantee	1 year from the date of purchase, when properly stored and handled.

► CRISPR RIBONUCLEOPROTEIN (RNP) + ssODN TRANSFECTION PROTOCOL

Fill in volumes below based on culture vessel used for transfection (Table 1).

A. Plate cells

1. Approximately 18-24 hours before transfection, plate cells in ___ml complete growth medium. Most adherent cell types should be $\sim 80\%$ confluent at the time of transfection.

For adherent cells: Plate cells at a density of $0.8\text{--}3.0 \times 10^5$ cells/ml.

For suspension cells: Plate cells at a density of $2.5\text{--}5.0 \times 10^5$ cells/ml.

2. Culture overnight.

B. Prepare *TransIT-X2[®]*:RNP complexes (Immediately before transfection)

1. Warm *TransIT-X2[®]* to room temperature and vortex gently before using.
2. Place ___ μl of OptiMEM[®] I Reduced-Serum Medium in a sterile tube.
3. Add ___ μl of a 50 μM guide RNA stock solution (12 nM final concentration per well). Mix gently by pipetting. NOTE: If using 2-part crRNA + tracrRNA, combine at a 1:1 molar ratio and incubate for 10 minutes at room temperature to anneal. Then add to tube containing OptiMEM[®].
4. Add ___ μl of a 30 μM Cas9 protein stock solution (6 nM final concentration per well). Mix gently by pipetting.
5. Incubate at room temperature for 10 minutes.
6. Add ___ μl ssODN (12 nM final concentration per well) to tube containing RNP mixture.
6. Immediately add ___ μl *TransIT-X2[®]* to the RNP:ssODN mixture. Mix gently by pipetting.
7. Incubate at room temperature for 15 minutes.

C. Distribute complexes to cells

1. Add the *TransIT-X2[®]*:RNP:ssODN complexes (prepared in Step B) drop-wise to different areas of the well. Gently rock plate for even distribution of complexes.
2. Incubate 24-72 hours.
3. Harvest cells and assay as required.

Table 1. Recommended starting conditions

Culture vessel	24-well plate	12-well plate	6-well plate
Surface area	1.9 cm ²	3.8 cm ²	9.6 cm ²
Complete growth medium	0.5 ml	1 ml	2.5 ml
Serum-free medium	50 μl	100 μl	250 μl
guide RNA (50 μM stock, 12 nM final in well)	0.12 μl	0.24 μl	0.6 μl
Cas9 Protein (30 μM stock, 6 nM final in well)	0.1 μl	0.2 μl	0.5 μl
ssODN (50 μM stock, 24 nM final in well)	0.12 μl	0.24 μl	0.6 μl
<i>TransIT-X2[®]</i> Reagent	1 μl	2 μl	5 μl

► Transfection Optimization:

The 2:1:2 ratio of guide RNA to Cas9 protein to ssODN (12 nM gRNA:6 nM Cas9:12 nM ssODN; final concentration per well) used in this protocol is a starting point for RNP + DNA oligo transfection. Further ratio optimization may be required for some cell types.

For more on transfection optimization, see the *TransIT-X2[®] full protocol (PDF)* or *Tips from the Bench*.

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