

# ***TransIT-X2<sup>®</sup>* Dynamic Delivery System for CRISPR/Cas9 Plasmid and gRNA Delivery**



Instructions for use with MIR 6000, 6003, 6004, 6005, 6006, 6010

## **SPECIFICATIONS**

<b>Storage</b>	Store <i>TransIT-X2<sup>®</sup></i> Dynamic Delivery System tightly capped at $-20^{\circ}\text{C}$ . <b>Before each use</b> , warm to room temperature and vortex gently.
<b>Product Guarantee</b>	1 year from the date of purchase, when properly stored and handled.

### **► CAS9 DNA + gRNA TRANSFECTION PROTOCOL**

#### **Fill in volumes below based on culture vessel used for transfection (Table 1).**

##### **A. Plate cells**

1. Approximately 18-24 hours before transfection, plate cells in \_\_\_ml complete growth medium. Most cell types should be  $\sim 80\%$  confluent at the time of transfection.

**For adherent cells:** Plate cells at a density of  $0.8\text{--}3.0 \times 10^5$  cells/ml.

**For suspension cells:** Plate cells at a density of  $2.5\text{--}5.0 \times 10^5$  cells/ml.

2. Culture cells overnight.

##### **B. Prepare *TransIT-X2<sup>®</sup>*:DNA:gRNA complexes (Immediately before transfection)**

1. Warm *TransIT-X2<sup>®</sup>* to room temperature and vortex gently before using.
2. Place \_\_\_ $\mu\text{l}$  of OptiMEM<sup>®</sup> I Reduced-Serum Medium in a sterile tube.
3. Add \_\_\_ $\mu\text{l}$  of a 50  $\mu\text{M}$  guide RNA stock solution (50 nM final concentration per well). Mix gently by pipetting. NOTE: If using 2-part crRNA + tracrRNA, first combine at a 1:1 molar ratio and incubate for 10 minutes at room temperature to anneal. Then add to tube containing OptiMEM<sup>®</sup>.
4. Add \_\_\_ $\mu\text{l}$  of plasmid DNA encoding Cas9. Mix gently by pipetting.
5. Add \_\_\_ $\mu\text{l}$  *TransIT-X2<sup>®</sup>* to the diluted DNA/gRNA mixture. Mix gently by pipetting.
6. Incubate at room temperature for 15-30 minutes.

##### **C. Distribute complexes to cells**

1. Add the *TransIT-X2<sup>®</sup>*:DNA:gRNA complexes (prepared in Step B) drop-wise to different areas of the well.
2. Gently rock plate for even distribution of complexes.
3. Incubate 24-72 hours.
4. Harvest cells and assay as required.

**Table 1.** Recommended starting conditions

Culture vessel	24-well plate	12-well plate	6-well plate
<b>Surface area</b>	1.9 cm <sup>2</sup>	3.8 cm <sup>2</sup>	9.6 cm <sup>2</sup>
<b>Complete growth medium</b>	0.5 ml	1 ml	2.5 ml
<b>Serum-free medium</b>	50 $\mu\text{l}$	100 $\mu\text{l}$	250 $\mu\text{l}$
<b>guide RNA (50 <math>\mu\text{M}</math> stock, 50 nM final)</b>	0.5 $\mu\text{l}$	1 $\mu\text{l}$	2.5 $\mu\text{l}$
<b>Cas9 Plasmid DNA (1 mg/ml stock)</b>	0.5 $\mu\text{l}$	1 $\mu\text{l}$	2.5 $\mu\text{l}$
<b><i>TransIT-X2<sup>®</sup></i> Reagent</b>	1 $\mu\text{l}$	2 $\mu\text{l}$	5 $\mu\text{l}$

#### **► Transfection Optimization:**

Determine the best *TransIT-X2<sup>®</sup>*:DNA ratio for each cell type. Start with 2  $\mu\text{l}$  of *TransIT-X2<sup>®</sup>* per 1  $\mu\text{g}$  of DNA. Vary the concentration of *TransIT-X2<sup>®</sup>* from 2-6  $\mu\text{l}$  per 1  $\mu\text{g}$  total DNA to find the optimal ratio.

For more on transfection optimization, see the *TransIT-X2<sup>®</sup>* [full protocol \(PDF\)](#) or [Tips from the Bench](#). Cell-type-specific recommendations are available at **Reagent Agent:** [mirusbio.com/ra](http://mirusbio.com/ra)

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## Reagent Agent<sup>®</sup>

Reagent Agent<sup>®</sup> is an online tool designed to help determine the best solution for nucleic acid delivery based on in-house data, customer feedback and citations.

Learn more at: [mirusbio.com/ra](http://mirusbio.com/ra)  
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