

# **TransIT®-mRNA Transfection Kit for CRISPR/Cas9** **mRNA and gRNA Delivery**



Instructions for use with MIR 2225, 2250, 2251, 2255, 2256

## **SPECIFICATIONS**

<b>Storage</b>	Store both <i>TransIT</i> ®-mRNA Reagent and mRNA Boost Reagent tightly capped at $-20^{\circ}\text{C}$ . <b>Before each use</b> , warm to room temperature and vortex gently.
<b>Product Guarantee</b>	1 year from the date of purchase, when properly stored and handled.

### **► CAS9 mRNA + gRNA TRANSFECTION PROTOCOL**

#### **Fill in volumes below based on culture vessel used for transfection (Table 1).**

##### **A. Plate cells**

1. Approximately 18-24 hours before transfection, plate cells in \_\_\_ml complete growth medium (per well). Most cell types should be  $\sim 80\%$  confluent on day of transfection.

**For adherent cells:** Plate cells at a density of  $0.8\text{--}3.0 \times 10^5$  cells/ml.

**For suspension cells:** Plate cells at a density of  $2.5\text{--}5.0 \times 10^5$  cells/ml.

2. Culture cells overnight.

##### **B. Prepare *TransIT*®-mRNA Reagent:mRNA Boost:RNA complexes**

1. Warm *TransIT*®-mRNA and Boost Reagent to room temperature and vortex gently.
2. Place \_\_\_ $\mu\text{l}$  of OptiMEM® I Reduced-Serum Medium in a sterile tube.
3. Add \_\_\_ $\mu\text{l}$  of guide RNA (50  $\mu\text{M}$  stock solution; 50 nM final concentration per well). Mix gently by pipetting. NOTE: If using 2-part crRNA + tracrRNA, first combine at a 1:1 molar ratio and incubate for 10 minutes at room temperature to anneal. Then add to tube containing OptiMEM®.
4. Add \_\_\_ $\mu\text{l}$  of mRNA encoding Cas9. Mix gently by pipetting.
5. Add \_\_\_ $\mu\text{l}$  of mRNA Boost Reagent. Mix gently by pipetting.
5. Add \_\_\_ $\mu\text{l}$  *TransIT*®-mRNA Transfection Reagent. Mix gently by pipetting.
6. Incubate at room temperature for 2-5 minutes to allow complexes to form.

##### **C. Distribute transfection complexes to cells**

1. Add the complexes (prepared in Step B) drop-wise to different areas of the wells.
2. Gently rock the culture vessel to evenly distribute the complexes.
3. Incubate 24-72 hours. NOTE: A post-transfection media exchange is not necessary.
4. Harvest cells and assay as required.

**Table 1.** Recommended starting conditions

Culture vessel	24-well plate	12-well plate	6-well plate
Surface area	1.9 cm <sup>2</sup>	3.8 cm <sup>2</sup>	9.6 cm <sup>2</sup>
Complete growth medium	0.5 ml	1 ml	2.5 ml
Serum-free medium	50 $\mu\text{l}$	100 $\mu\text{l}$	250 $\mu\text{l}$
guide RNA (50 $\mu\text{M}$ stock, 50 nM final)	0.5 $\mu\text{l}$	1 $\mu\text{l}$	2.5 $\mu\text{l}$
Cas9 mRNA (1 mg/ml stock)	0.5 $\mu\text{l}$	1 $\mu\text{l}$	2.5 $\mu\text{l}$
mRNA Boost Reagent	0.5 $\mu\text{l}$	1 $\mu\text{l}$	2.5 $\mu\text{l}$
<i>TransIT</i> ®-mRNA Transfection Reagent	0.5 $\mu\text{l}$	1 $\mu\text{l}$	2.5 $\mu\text{l}$

#### **► Transfection Optimization:**

Cell type, cell confluency, reagent volume, and post-transfection incubation time are a few of the key parameters that affect the outcome of transfection experiments. For more on transfection optimization, see the *TransIT*®-mRNA [full protocol \(PDF\)](#) or [Tips from the Bench](#).

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