

TransIT®-Keratinocyte Transfection Reagent

Protocol for MIR 2800, 2804, 2805, 2806

Quick Reference Protocol, SDS and Certificate of Analysis available at mirusbio.com/literature

mirus bio®

INTRODUCTION

TransIT®-Keratinocyte Transfection Reagent is specifically optimized to provide exceptional transfection efficiency of plasmid DNA in primary and immortalized keratinocytes. TransIT-Keratinocyte Reagent provides all the attributes of the trusted TransIT series of transfection reagents: high transfection efficiency, low toxicity, serum compatibility, simplicity of use and reproducibility. Transfection with TransIT Keratinocyte Reagent does not require medium changes and can be carried out in serum-containing medium. TransIT-Keratinocyte is suitable for both transient and stable transfection.

SPECIFICATIONS

Storage	Store TransIT-Keratinocyte Reagent at 4°C. Before each use , warm to room temperature and vortex gently.
Product Guarantee	1 year from the date of purchase, when properly stored and handled.



Warm TransIT-Keratinocyte to room temperature and vortex gently before each use.

MATERIALS

Materials Supplied

TransIT-Keratinocyte Transfection Reagent is supplied in **one** of the following formats.

Product No.	Quantity
MIR 2804	1 × 0.4 ml
MIR 2800	1 × 1.0 ml
MIR 2805	5 × 1.0 ml
MIR 2806	10 × 1.0 ml

Materials required, but not supplied

- Cultured cells
- Appropriate cell culture medium
- Purified plasmid DNA
- Serum-free medium (e.g. Opti-MEM® I Reduced-Serum Medium)
- Sterile tube for transfection complex preparation
- Micropipets
- Reporter assay as required
- *Optional:* Selection antibiotic (e.g., G418 or Hygromycin B) for stable transfection

For Research Use Only

BEFORE YOU START:

Important Tips for Optimal Plasmid DNA Transfection

Optimize reaction conditions for each keratinocyte cell type to ensure successful transfections. The suggestions below yield high efficiency transfection of most keratinocyte cell types using TransIT-Keratinocyte Transfection Reagent. **Table 1** presents recommended starting conditions depending on culture vessel size.

- **Cell density (% confluence) at transfection.** Determine the optimal cell density for each keratinocyte subtype to maximize transfection efficiency. Divide the cells 18–24 hours before transfection to ensure that the cells are actively dividing and reach the appropriate cell density (generally $\geq 80\%$ confluence) at the time of transfection.
- **DNA purity.** Use highly purified, sterile, and contaminant-free DNA for transfection. Plasmid DNA preparations that are endotoxin-free and have $A_{260/280}$ absorbance ratio of 1.8–2.0 are desirable. DNA prepared using miniprep kits is not recommended as it might contain high levels of endotoxin. We recommend using MiraCLEAN® Endotoxin Removal Kit (MIR 5900) to remove endotoxin from your DNA preparation.
- **Ratio of TransIT-Keratinocyte Reagent to DNA.** Determine the best TransIT-Keratinocyte Reagent:DNA ratio for each keratinocyte cell type. Start with 3 μl of TransIT-Keratinocyte Reagent per 1 μg of DNA. Vary the concentration of TransIT-Keratinocyte Reagent from 2–8 μl per 1 μg DNA to find the optimal ratio. **Table 1** provides recommended starting conditions based on cell culture vessel size.
- **Complex formation conditions.** Prepare TransIT-Keratinocyte Reagent:DNA complexes in serum-free growth medium. Mirus recommends Opti-MEM I Reduced-Serum Medium.
- **Cell culture conditions:** Culture cells in the appropriate medium, with or without serum. There is no need to perform a medium change to remove the transfection complexes.
- **Presence of antibiotics:** Antibiotics may inhibit transfection complex formation and therefore should be excluded from the complex formation step. Transfection complexes can be added directly to cells grown in complete culture medium containing serum and low levels of antibiotics (0.1–1X final concentration of penicillin/streptomycin mixture).
- **Post-transfection incubation time.** Determine the best incubation time post-transfection for each keratinocyte cell type. The optimal incubation time is generally 24–72 hours, but will vary depending on the goal of the experiment, nature of the plasmid, and the half-life of the expressed protein.

Table 1. Recommended starting conditions for DNA transfections with TransIT-Keratinocyte Transfection Reagent.

Culture vessel	96-well plate	48-well plate	24-well plate	12-well plate	6-well plate	10-cm dish	T75 flask
Surface area	0.35 cm^2	1.0 cm^2	1.9 cm^2	3.8 cm^2	9.6 cm^2	59 cm^2	75 cm^2
Complete growth medium	92 μl	263 μl	0.5 ml	1.0 ml	2.5 ml	15.5 ml	19.7 ml
Serum-free medium	9 μl	26 μl	50 μl	100 μl	250 μl	1.5 ml	1.9 ml
DNA (1 $\mu\text{g}/\mu\text{l}$ stock)	0.1 μl	0.25 μl	0.5 μl	1 μl	2.5 μl	15 μl	19 μl
TransIT-Keratinocyte Reagent	0.3 μl	0.75 μl	1.5 μl	3 μl	7.5 μl	45 μl	57 μl



Do not use DNA prepared using miniprep kits for transfection.



Do not use serum or antibiotics in the medium during transfection complex formation.



Surface areas are based on Greiner tissue culture plates and Falcon 10-cm dishes and T75 flasks. All volumes given are per well (or per dish) for a given culture vessel.

If small volumes of TransIT-Keratinocyte need to be pipetted, dilute the reagent in serum-free medium before each use to avoid pipetting errors. **Do not** store diluted TransIT-Keratinocyte Reagent.

PLASMID DNA TRANSFECTION PROTOCOL

The following procedure describes how to perform plasmid DNA transfections in 6-well plates. The surface areas of other culture vessels are different and transfections must be scaled accordingly. Appropriately increase or decrease the amounts of serum free medium, TransIT-Keratinocyte Reagent, DNA and complete culture medium based on the surface area of the cell culture vessel (please refer to **Table 1** on Page 2).

Transient plasmid DNA transfection protocol per well of a 6-well plate



Divide cultured cells 18–24 hours before transfection to ensure active cell division at the time of transfection.

A. Plate cells

1. Approximately 18–24 hours before transfection, plate cells in 2.5 ml complete growth medium per well in a 6-well plate. Ideally cells should be $\geq 80\%$ confluent prior to transfection.
2. Incubate cell cultures overnight.

B. Prepare TransIT-Keratinocyte Reagent:DNA complex

(Immediately before transfection)

1. Warm TransIT-Keratinocyte Reagent to room temperature and vortex gently before using.
2. Place 250 μ l of Opti-MEM I Reduced-Serum Medium in a sterile tube.
3. Add 2.5 μ g (2.5 μ l of a 1 μ g/ μ l stock) plasmid DNA. Pipet gently to mix completely.
4. Add 7.5 μ l TransIT-Keratinocyte Reagent to the diluted DNA mixture. Pipet gently to mix completely.
5. Incubate at room temperature for 15–30 minutes to allow sufficient time for complexes to form.



There is no need to change culture medium after transfection. If required, perform a medium change at least 4 hours post-transfection.

C. Distribute the complexes to cells in complete growth medium

1. Add the TransIT-Keratinocyte Reagent:DNA complexes drop-wise to different areas of the wells.
2. Gently rock the culture vessel back-and-forth and from side-to-side to evenly distribute the TransIT-Keratinocyte Reagent:DNA complexes.
3. Incubate for 24–72 hours. It is not necessary to replace the complete growth medium with fresh medium.
4. Harvest cells and assay as required.

For generating stable cell transfectants, passage the cells 48–72 hours post-transfection in complete growth medium containing the appropriate selection antibiotic such as G418 or Hygromycin B. Maintain selection for 1–2 weeks, allowing selection of cells that have undergone stable integration of DNA.

TROUBLESHOOTING GUIDE

Problem	Solution
LOW PLASMID DNA TRANSFECTION EFFICIENCY	
<i>TransIT</i> -Keratinocyte Reagent was not mixed properly	Warm <i>TransIT</i> -Keratinocyte to room temperature and vortex gently before each use.
Suboptimal <i>TransIT</i> -Keratinocyte Reagent:DNA ratio	Determine the best <i>TransIT</i> -Keratinocyte Reagent:DNA ratio for each keratinocyte cell type. Titrate the <i>TransIT</i> -Keratinocyte Reagent from 2–8 µl per 1 µg DNA. Refer to “Before You Start” on Page 2.
Suboptimal DNA concentration	Confirm DNA concentration and purity. Use plasmid DNA preps that have an A _{260/280} absorbance ratio of 1.8–2.0.
Low-quality plasmid DNA	The optimal DNA concentration generally ranges between 1–3 µg/well of a 6-well plate. Start with 2.5 µg/well of a 6-well plate. Consider testing more or less DNA while scaling the amount of <i>TransIT</i> -Keratinocyte Transfection Reagent accordingly.
Inhibitor present during transfection	Use highly purified, sterile, endotoxin and contaminant-free DNA for transfection. We recommend using Mirus Bio’s MiraCLEAN Endotoxin Removal Kit (MIR 5900) for removal of endotoxin from your DNA preparation. Alternatively, use cesium chloride gradient or anion exchange purified DNA which contains levels of endotoxin that do not harm most cells.
Incorrect vector sequence	Do not use DNA prepared using miniprep kits as it may contain high levels of endotoxin.
Transfection incubation time	Serum and antibiotics inhibit transfection complex formation. Prepare <i>TransIT</i> -Keratinocyte Reagent:DNA complexes in serum-free growth medium. We recommend Opti-MEM I Reduced-Serum Medium. Once transfection complexes are formed, they can be added directly to cells cultured in complete growth medium containing serum and 0.1–1X antibiotics.
Cells not actively dividing at the time of transfection	Divide the culture at least 18–24 hours before transfection to ensure that the cells are actively dividing and reach optimal cell density at time of transfection.
Precipitate formation during transfection complex formation	During complex formation, scale all reagents according to Table 1 on page 2 including serum-free medium, <i>TransIT</i> -Keratinocyte and plasmid DNA. Precipitation may be observed when excess DNA is used during complex formation. This may negatively impact transfection efficiency. To avoid precipitation when using high concentrations of DNA, increase the volume of serum-free medium during complex formation by two-fold.
Proper experimental controls were not included	To verify efficient transfection, use <i>TransIT</i> -Keratinocyte Reagent to deliver a positive control such as a luciferase, beta-galactosidase or green fluorescent protein (GFP) encoding plasmid. To assess delivery efficiency of plasmid DNA, use Mirus’ <i>Label IT</i> ® Nucleic Acid Labeling Kits to label the target plasmid or Mirus’ prelabeled <i>Label IT</i> Plasmid Delivery Controls (please refer to Related Products on Page 6).

TROUBLESHOOTING GUIDE continued

Problem	Solution
HIGH CELLULAR TOXICITY	
Transfection complexes and cells not mixed thoroughly after complex addition	Add <i>TransIT</i> -Keratinocyte Reagent:DNA complexes drop-wise to different areas of the wells containing cells. Gently rock the dish back-and-forth and from side-to-side to distribute the complexes evenly. Do not swirl or rotate the dish, as this may cause uneven distribution.
Transfection complexes added to cells cultured in serum-free medium	Allow <i>TransIT</i> -Keratinocyte Reagent:DNA complexes to form in serum-free medium, then add these complexes to cells cultured in complete growth medium. The presence of serum in the growth medium improves transfection efficiency and reduces cytotoxicity. No culture medium change is required after the addition of transfection complexes to cells.
	Use highly purified, sterile, endotoxin and contaminant-free DNA for transfection.
Endotoxin-contaminated plasmid DNA	We recommend using Mirus Bio's MiraCLEAN Endotoxin Removal Kit (MIR 5900) for removal of any traces of endotoxin from your DNA preparation. Alternatively, use cesium chloride gradient or anion exchange purified DNA which contains levels of endotoxin that do not harm most cells.
	Do not use DNA prepared using miniprep kits as it may contain high levels of endotoxin.
Expressed target gene is toxic to cells	Compare toxicity levels against a cells alone control and cells transfected with an empty vector to assess the cytotoxic effects of the target protein being expressed.
	If lower levels of target gene expression are desired in your transfection experiments, consider reducing the amount of target plasmid. Maintain the optimal <i>TransIT</i> -Keratinocyte:DNA ratio by using carrier DNA such as an empty cloning vector.
Cell density not optimal at time of transfection	Determine the best cell density for each keratinocyte cell type to maximize transfection efficiency. Use this cell density in subsequent experiments to ensure reproducibility. For most cell types, $\geq 80\%$ confluence is recommended at transfection, but use of higher or lower densities may increase cell viability depending on cell type.
Cell morphology has changed	Mycoplasma contamination can alter cell morphology and affect transfection efficiency. Check your cells for Mycoplasma contamination. Use a fresh frozen stock of cells or use appropriate antibiotics to eliminate Mycoplasma.
	A high or low cell passage number can make cells more sensitive and refractory to transfection. Maintain a similar passage number between experiments to ensure reproducibility.

RELATED PRODUCTS

- Ingenio® Electroporation Solution and Kits
- *Label IT*® Plasmid Delivery Controls
- *Label IT*® Nucleic Acid Labeling Kits
- MiraCLEAN® Endotoxin Removal Kits
- Cell Culture Antibiotic Solutions
- *TransIT-X2*® Dynamic Delivery System
- *TransIT*®-2020 Transfection Reagent
- *TransIT-PRO*® Transfection Kit
- *TransIT*®-LT1 Transfection Reagent
- *TransIT*® Cell Line Specific Transfection Reagents and Kits



Reagent Agent®

Reagent Agent® is an online tool designed to help determine the best solution for nucleic acid delivery based on in-house data, customer feedback and citations.

Learn more at:
www.mirusbio.com/ra

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